



Original Research Article

LIPOSOME SYNTHESIS AND CHARACTERIZATION IN THE BORON NEUTRON CAPTURE THERAPY OF CANCER

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Abstract: BNCT, or boron neutron capture therapy, is a cancer treatment that uses neutron irradiation to kill cancer cells that have taken up ¹⁰B target chemicals specifically. There are short-range particles that may cause cell harm when they are captured by neutrons and ¹⁰B atoms. BNCT, on the other way, is meant to target cancer cells while preserving normal ones. BNCT's success is dependent on the boron carrier utilised. MAC and Na₃[ae-B₂₀H₁₇NH₃] (TAC) are incorporated into the bilayer membrane of these liposomes, while the hydrophilic species Na₃[ae-B₂₀H₁₇NH₃] (TAC) is contained in this aqueous core of the liposomes. Liposomes having a diameters of 83 nm were injected intravenously into hamsters. To compare tumour boron concentrations to precancerous tissue levels, the ratio was 10:1 after 48 hours, with the tumour boron concentration being 67 ppm and precancerous tissue being 11 ppm. Precancerous tissue received a 5-Gy dosage of neutron irradiation (equivalent to 21 Gy in tumour), which result in a 70% overall tumour responses (OR) after a 4-week follow-up periods. The OR rate for the beam-only procedure was just 28%. At an interval of 4, 6, or 8 weeks, BNCT therapy with liposome re-administration resulted in OR rates of 70–88 percent, with a full response of 37–52 percent. We could prolong follow-up to 16 weeks following the first therapy because of the positive therapeutic response. Normal tissue was shown to be unaffected by radiation. There was only minor mucositis in dose-limiting precancerous tissue with a tumour response of 70–88 percent with these liposomes.

Keywords: Boron neutron capture therapy, liposomes, tumour, therapeutic response.

INTRODUCTION

Liposomes have proven to be an effective pharmacological treatment for cancer. Although liposome uses in cancer therapies have been studied in great detail and merit a thorough evaluation, this is outside the purview of this study and review. Nonetheless, this article discusses the most effective uses of liposomes in cancer treatment. It has been demonstrated that a variety of liposomal formulations of anti-cancer medications can deliver the medications to these solid cancer cell locations with fewer toxicities than the free drug quantity. As of right now, there are numerous formulations available on the market and in clinical trials for use as drug delivery vehicles for chemotherapy [1-5]. The first liposomal preparation to be licensed by the FDA for

the treatment of Kaposi's sarcoma cells in HIV patients was Doxil, a PEGylated liposomal formulation. Johnson & Johnson sells PEGylated liposomal formulations called Doxil (US) or Caelyx (outside the US) that encapsulate the anticancer medication doxorubicin. An imbalance between Doxil's supply and demand was noted in 2011 when manufacturing facilities were briefly closed because of problems with quality control. Liposome have rapidly spread into the various science and application and well over 10 Thousand research article have jet been publish. Their mechanical and surfaceS property are defined by the compositions, size and lamellarity, where stability also play an important roles, which depend additional on external factor, such as temperatures, pressure, physical

stress, chemical and biological agent, and the presences of various cell type. Liposome are spherical, self-closed structure composes of curved lipid bilayer which encapsulate parts of the solvents, in which they float. They are unilamellar (consist of one concentric membrane) or multilamellar (consist of several concentric membrane).[10-12]

2. MATERIALS & METHODS

When it comes to developing novel formulations, preformulation studies are critical because they provide researchers a tool for selecting desirable excipients that are also compatible with the medicine under consideration. Particle sizes analysis, XRD analyses, DSC analyses, solubility analyses, partition coefficient, solution stability, and compatibility investigations are many critical preformulation studies. All of the above listed qualities have a substantial impact on the formulation development process. Liposome size is crucial, for example, to prevent the first pass effect. As a result, all excipients should be chosen such that their influence on liposomal size is minimised. Particle size measurement of excipients is thus critical in the design of the liposome manufacturing process. Further sonication is used to shrink the liposome until the particle size is significantly decreased.[13]

UV spectroscopy is an effective instrument for determining the quality and quantity of any given medicine. Using UV spectroscopy, the solubility and dissolution rate may be determined in advance of formulation. Using UV spectroscopy, researchers are able to conduct several stability tests. HPLC typically makes use of UV detectors. As a result, UV spectroscopy is critical in the early phases of preformulation (because to its high accuracy and simplicity).[14]

IR spectroscopy is commonly used to identify a medication molecule by its unique fingerprint identification pattern. Pharmaceutical solids may be physically characterized using FTIR spectroscopy, a method that uses a Fourier transform to change the infrared spectrum. Drug-excipient compatibility studies and the determination of polymorphism of solid crystals may both be carried out using infrared spectroscopy (IR) during preformulation.[15]

Atomic structure may be determined using the X-ray diffractometry method. Using this method, crystalline materials may be distinguished from noncrystalline ones. Analyses performed using these methods are nondestructive; therefore, samples in any form may be used.

Drug-excipient compatibility is determined by differential scanning calorimetry (DSC) analysis. To see whether a medicine and its excipients were compatible, researchers used a controlled temperature increase and compared the resulting

thermograms to those of the individual components. If there is no interaction, the excipients may be used with the chosen medicinal molecule without concern. As many factors as heating rate, humidity, pan arrangement, and particle size influence the DSC study findings. PXRD and FTIR are examples of complementary methods that should be utilised to better understand the interactions and how they relate to formulation developments.

For liposome formulation, solubilities studies would be carried out on two the medication and the excipient in various polarity solvent. Liposome production is largely dependent on the drug's solubility. Liposomes typically have water compartments for hydrophilic medicinal molecules and a lipid bilayer for hydrophobic and amphiphilic chemical carriers. The nature of the medicine is thus crucial for liposome formulation development and stability.

Another critical preformulation parameters is the partitions coefficient. The hydrophilic-lipophilic balance of a pharmacological substance is shown by the octanol-water partition ($K_{o/w}$) coefficient. This log value ratio is referred to as log P value and is used to estimate a drug's permeability through a biological membrane. The amount of drug entrapped in a liposomal formulation is determined by the drug's partition coefficient. Partition coefficient, composition of the lipid bilayer, and drug molecule size all have a role in drug delivery. Various preparation medium should be tested for solution stability before using them to prepare medicines. Because throughout the production process, the medicine is often in touch with various solvents and, as a result, comes into contact with bodily fluids. If a medication degrades in any way, the formulation will fail to provide the intended therapeutic result.

The compatibility research is a critical part of the preformulation process since it informs the selection of excipients and packaging components. There should be great attention used while selecting drugs and excipients for therapeutic response from any dose form.

The research of drug-excipient interactions is crucial for the creation of drug-loaded liposomes. A drug-to-excipient compatibility study is required to identify any interactions that may occur during the production process or storage. It's possible that this sort of interaction may lead to product deterioration or impurities in the final dose. Toxic impurities might have undesirable side effects.[16-18]

RESULTS AND DISCUSSION

EFFECT OF CHOLESTEROL CONTENT ON THE PHYSICO-CHEMICAL PROPERTIES OF LIPOSOMES

Particle size, percent entrapment efficiency, and other physicochemical parameters were tuned for many batches of liposomal formulations generated using the REV technique (percent EE). We experimented with different cholesterol doses. The quantity of cholesterol utilized had a considerable effect on the size of the liposomal formulations' vesicles. In addition to this, the EE percentage dropped for liposomal formulations with higher cholesterol content.. The ratio of 5-FU to lipid mass was maintained at 1:10 at all times. A 5:1 to a 5:5 SPC molar ratio was used (SPC:CHOL). All subsequent trials used the same formulation, PREV-1 photosensitive liposomal, as the first. Figures 5.2 and 5.3 provide an example of a vesicle size study of manufactured liposomes.

Table 1: Effect of cholesterol content on conventional liposomal formulations

Formulation code	PC	Chol	Mean vesicle size (nm)	%EE
REV-I	6	2	173.9 ± 7.9	59.5 ± 2.8
REV-II	6	3	263.0 ± 7.9	50.6 ± 2.2
REV-III	6	4	185.0 ± 6.8	40.3 ± 4.3
REV-IV	6	5	247.0 ± 7.6	33.4 ± 4.9
REV-V	6	6	196.2 ± 8.4	30.7 ± 3.4

All data are expressed as the means ± standard deviation ($n = 3$).

Fig. 1: Representative vesicle size analysis of 5:1 conventional liposomal formulation

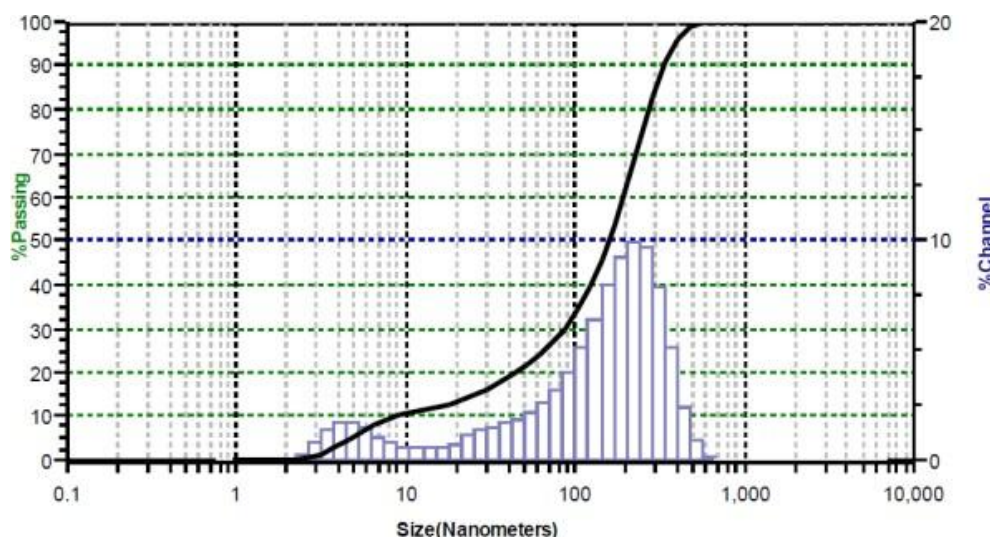


Table 2: Effect of cholesterol content on photosensitive liposomal formulations

Formulation code	PC	Chol	Mean vesicle size (nm)	%EE
PRV-1	6	2	121.0 ± 5.8	45.4 ± 3.5
PRV-2	6	3	132.8 ± 9.7	37.5 ± 2.8
PRV-3	6	4	299.6 ± 4.9	27.0 ± 3.5
PRV-4	6	5	307.8 ± 4.8	18.1 ± 3.2
PRV-5	6	6	318.5 ± 7.0	12.4 ± 1.4

All data are expressed as the means ± standard deviation ($n = 3$).

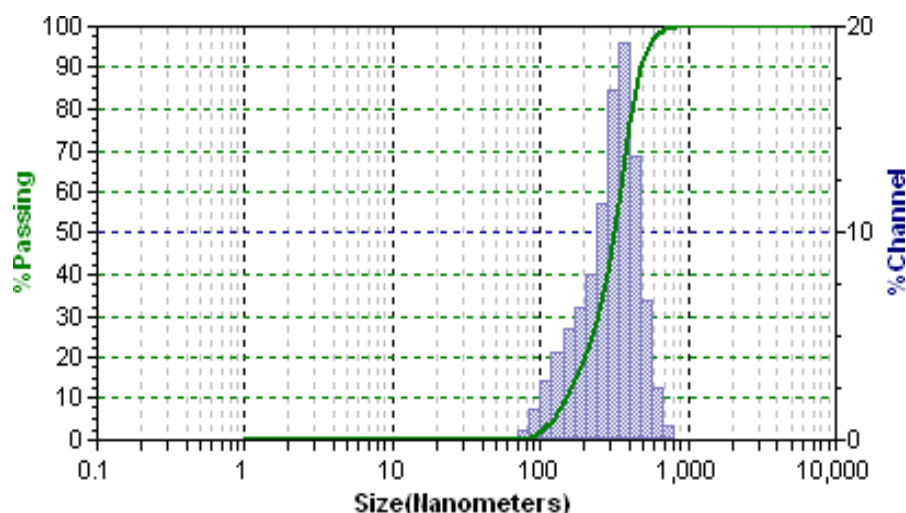


Fig.2:Representative vesicle size analysis of 5:1 photosensitive liposomal formulation

The size of the vesicle and its dispersion are critical factors in predicting medication release at the target location. Modifications and controls should be implemented during preparation in such a way that the in vivo or ex vivo performance may be predicted. There are several preparation methods that may be used to provide a homogeneous formulation with an optimal size range. In this respect, the REV method offers an alternative. The current investigation found that when the molar cholesterol ratio increased from 5:1 to 5:5, the size of the vesicle increased in both conventional liposomes and photosensitive liposomes (SPC:CHOL). To conclude, researchers found that when cholesterol concentration increased, so did the ability of bilayer vesicles to form more rigidly, according to Shivare et al.

When cholesterol content was increased from a molar ratio of 5:1 to 5:5, the percent EE decreases (Kulkarni, Betageri, and Singh 229; Ramana et al. 57). The involvement of cholesterol in liposome production is linked to its act of accumulation in phospholipid bilayer structural cavities, which eventually improve stiffness and packing density. Due to cholesterol's space-filling properties, it reduces the loading of hydrophilic medicines.

In vitro drug release from liposomal formulations

In vitro release studies are less costly than testing in vivo performance of various formulations in terms of cost efficiency. Photosensitive liposomes and regular liposomes were both used in this investigation to see how UV light affected medication release. A UV Spectrophotometer was used to measure the quantity of 5-FU that had been released (Shimadzu, Japan, Model-2460). The dialysis tube technique was used to characterise formulations for in vitro 5-FU release. This experiment employed a liposomal formulation that had not been exposed to sunlight as a control. As a result, it was found that photosensitive liposomes released 100 percent of the medicine they contained after 15 hours, but regular liposomes only released 67 percent. The formulations were released in zero-order sequence after Cirli and Hasirci (85–96).

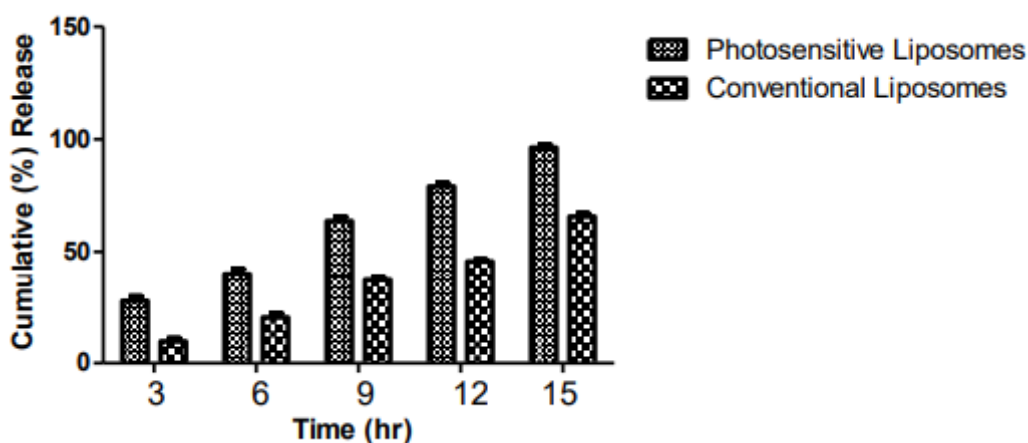


Fig. 3: Cumulative % drug (5-FU) release from different liposomal formulation

Kinetic analysis of drug releases

When using innovative drug delivery systems such as liposomes, it is critical to know when the medication will be released from the system. The expected biological impact of a medicine is totally dependent on its release kinetics from nanocarriers. When it comes to forecasting the impact of formulation parameters on drug release, many kinetic models have proven quite important. The transfer of drug molecules from donor liposomes to acceptor liposomes has been explained by two separate methods. Different *in vitro* kinetic models including zero-order, first-order, and Korsmeyer-peppas models equations were handled for varied drug release data in the current investigation. As a result, drug release mechanisms from produced formulations may be better predicted.

Most correlation coefficients were used to collect and evaluate the data. Table 5.3 summarises the correlation coefficients (R) derived from several kinetic models.

There was zero order kinetics in the photosensitive liposomal formulation. Using zero-order equations, researchers were able to accurately describe the *in vitro* release profile of the photosensitive liposomal formulations medications.

CONCLUSION

Every one of the conjugates tested displayed a strong affinity for their receptors, were internalised, and were stable in the cell after intracellular retention. Internalization is critical for BNCT because the chemical is more harmful if decay occurs within the cell rather than on the cellular membranes due to the limited range of the particles. The displacement test was used to determine the specificity, and the results reveal unspecific binds ranging from the 10% to 30%. Similar findings have been made by other teams. Free folate added in excess by Gabizon et al. blocked 86% of the binds of constructs including folate-PEG-liposomes. According to Lee et al., folate liposomes and a folic acid block had a 30% unspecific binding rate (34). There was a 50% decrease in absorption after adding free anti-CD19 antibody as opposed to taking anti-CD19- liposomes, according to Lopez de Menezes et al (46). To measure uptake in healthy tissue, the unspecific uptake must be examined in an *in vivo* setting. Unspecific absorption is less of a concern in BNCT since only the irradiated regions are subjected to the harmful consequences.

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